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     Hif2 alpha, Ap-2 beta, and Et-1 cooperatively regulate
TI
development of the
     ductus arteriosus.
ΑU
     Ivey, Kathryn N. [Reprint Author]; Garg, Vidu; Garcia, Joseph;
Zhao, Feng;
     Gelb, Bruce D.; Srivastava, Deepak
     Univ Texas, SW Med Ctr, Dallas, TX 75230 USA
CS
     Circulation, (OCT 26 2004) Vol. 110, No. 17, Suppl. S, pp. 59.
SO
     Meeting Info.: 77th Scientific Meeting of the
American-Heart-Association.
     New Orleans, LA, USA. November 07 -10, 2004. Amer Heart Assoc.
     CODEN: CIRCAZ. ISSN: 0009-7322.
DT
     Conference; (Meeting)
     Conference; Abstract; (Meeting Abstract)
LA
     English
     Entered STN: 1 Dec 2005
ED
     Last Updated on STN: 1 Dec 2005
     Development of the ductus arteriosus, a fetal vessel bridging the
AB
     pulmonary and systemic vasculature, involves specification of
highly
     contractile, oxygen-responsive vascular smooth muscle.
of this
     developmental process results in patent ductus arteriosus, the
third most
     common congenital heart defect. We identified an individual
with patent
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ductus arteriosus carrying a heterozygous nonsense mutation in

the gene

encoding the endothelin-A receptor (ETA). Analysis in mouse embryos

revealed that although Et-A was expressed uniformly throughout the

vascular smooth muscle of the great vessels and ductus arteriosus, the

gene encoding the Et-A ligand, endothelin-1 (Et-1), was uniquely confined

to the smooth muscle cells of the ductus arteriosus at embryonic day  $(\mathsf{E})$ 

13.5. A hypoxic response element upstream of Et-1 is required for hypoxic

induction of Et-1 expression. We found that Hif2 alpha, encoding a bHLH/PAS domain-containing hypoxia-inducible transcription factor was specifically expressed in the smooth muscle of

the ductus arteriosus at E13.5 with sharp borders at the aortic and

pulmonary artery junctions. Although embryonic lethality of Hif2 alpha(-/-) embryos precluded analysis of its role in ductal development,

we examined mice lacking Tfap2 beta, a transcription factor associated

with patent ductus arteriosus in humans with Char syndrome. We found that

Tfap2 beta was required for expression of both Et-1 and Hif2 alpha in

smooth muscle, but not endothelial cells of the arterial duct. Histological analysis of Tfap2 beta(-/-) mouse embryos showed that

although smooth muscle cells were present in the ductus arteriosus, they

failed to maintain their highly differentiated state. Finally, Hif2 alpha

functioned as a negative regulator of Ap-2 beta-induced transcription,

suggesting a negative feedback loop that may refine the Ap-2 beta signal

during ductal development. The mechanism of negative regulation involved

Hif2 alpha disruption of sequence-specific DNA binding by Ap-2 beta and

was P300-independent. Our data, along with the requirement of AP-2 beta

for closure of the ductus arteriosus in humans, suggest that Hif2 alpha,

Ap-2 beta, and Et-1 cooperatively regulate development of the ductus

arteriosus.

L4 ANSWER 2 OF 2 MEDLINE on STN AN 2000026909 MEDLINE

DUPLICATE 1

DN PubMed ID: 10559262

TI EPAS1 trans-activation during hypoxia requires p42/p44 MAPK.

AU Conrad P W; Freeman T L; Beitner-Johnson D; Millhorn D E

CS University of Cincinnati, College of Medicine, Department of Molecular and

Cellular Physiology, Cincinnati, Ohio 45267-0576, USA.

NC HL07571 (NHLBI)

R37HL33831 (NHLBI)

RO1HL59945 (NHLBI)

SO The Journal of biological chemistry, (1999 Nov 19) Vol. 274, No. 47, pp.

33709-13.

Journal code: 2985121R. ISSN: 0021-9258.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

EM 199912

ED Entered STN: 20000113

Last Updated on STN: 20000113

Entered Medline: 19991214

AB Hypoxia is a common environmental stress that regulates gene expression

and cell function. A number of hypoxia-regulated transcription factors

have been identified and have been shown to play critical roles in

mediating cellular responses to hypoxia. One of these is the endothelial

PAS-domain protein 1 (EPAS1/HIF2-alpha/HLF/HRF). This
 protein is 48% homologous to hypoxia-inducible factor 1-alpha
 (HIF1-alpha). To date, virtually nothing is known about the
signaling

pathways that lead to either EPAS1 or HIF1-alpha activation. Here we show

that EPAS1 is phosphorylated when PC12 cells are exposed to hypoxia and

that p42/p44 MAPK is a critical mediator of EPAS1 activation. Pretreatment of PC12 cells with the MEK inhibitor, PD98059,

completely

blocked hypoxia-induced trans-activation of a hypoxia response element

(HRE) reporter gene by transfected EPAS1. Likewise, expression of a

constitutively active MEK1 mimicked the effects of hypoxia on HRE reporter

gene expression. However, pretreatment with PD98059 had no effect on

EPAS1 phosphorylation during hypoxia, suggesting that MAPK targets other

proteins that are critical for the trans-activation of EPAS1. We further

show that hypoxia-induced trans-activation of EPAS1 is independent of Ras.

Finally, pretreatment with calmodulin antagonists nearly completely

blocked both the hypoxia-induced phosphorylation of MAPK and the EPAS1

trans-activation of HRE-Luc. These results demonstrate that the MAPK

pathway is a critical mediator of EPAS1 activation and that activation of

MAPK and EPAS1 occurs through a calmodulin-sensitive pathway and not

through the GTPase, Ras. These results are the first to identify a  $\,$ 

specific signaling pathway involved in EPAS1 activation.